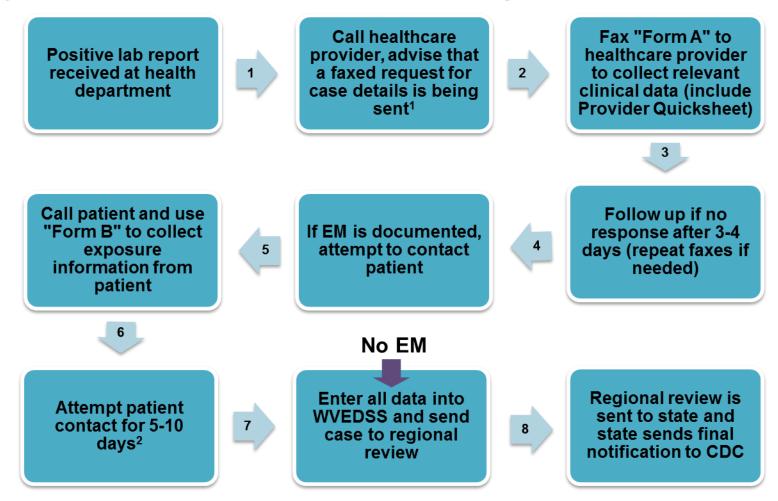


2017 LYME DISEASE CASE INVESTIGATION TOOLKIT

This toolkit can be used by local health department staff to facilitate Lyme disease case investigations. Items in the toolkit include:

- Case Investigation Flowchart
- Form A for Healthcare Providers
- Form B for Patients with Erythema Migrans (EM)
- Enzyme Immunoassay (EIA) Tips
- Interpretation IgM/IgG Western Blots
- 2017 Case Ascertainment Guide
- Provider Quicksheet
- 2017 Low and High Incidence Lyme Disease States

Lyme Disease Case Investigation Flowchart



¹ Request copies of any supplemental lab results; also ask for demographic data (e.g. race and ethnicity).

² Attempt to get in contact with patients through different methods. Try calling at least three times at different times of the day. Try alternate contact numbers and addresses. Mail a certified letter to the patient's address. Be sure to document all attempts.



Division of Infectious Disease Epidemiology

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Form A: Lyme Disease Assessment Tool (2017) For Healthcare Providers



Dear Healthcare Provider:							
The County Health Department has been notified of a positive Lyme disease laboratory report or patient (DOB:/). In order to comply with state and federal							
for patient infectious disease reporting req							
return this completed sheet via				s patient. Flease			
•	•						
A. Have you contacted this p				□YES □NO			
B. Date of first symptom ons				☐YES ☐NO			
C. Did this patient have an er If yes, where was the patient v	vhen he/she was likely bitten	ring <u>at least</u> 5 by an infected t	tick in the past 30 days?	□YES □NO			
(County):	(State):	of lote stage	Lyma disassa	□YES □NO			
D. Did patient exhibit any of t		or rate-stage	Lyme disease?	∐YES ∐NO			
Rheumatologic/musculoskeletal (mark one):							
Recurrent, brief attacks objective joint swelling (one or few joints)							
Chronic arthritis preceded by brief attacks (one or few joints)							
Other:							
No rheumatologic/musculoskeletal symptoms associated with LD were observed							
Neurologic (mark all that apply	<i>/</i>):						
Lymphocytic meningitis							
Radiculoneuropathy Encephalomyelitis Other:							
No neurologic symptoms associated with LD were observed							
	sociated with LD were obs	oci ved					
Cardiovascular (mark one):							
Acute onset of high-grade (2 nd or 3 rd degree) atrioventricular conduction defects (that resolves in days to weeks)							
Other:							
☐No cardiac symptoms assoc	iated with LD were observ	red					
• •				•			
E. Did you diagnose this patie	ent as having Lyme dise	ease?	□YES □NO)			
F. Please indicate what testing	g was ordered for this p	atient and an	y known results.				
Test Ordered	Date		Result				
Serology screen (IFA/EIA)	1 1		Negative Equivoca	I Pending			
Borrelia burgdorferi IgG WB	1 1	Positive _	Negative Pending				
Borrelia burgdorferi IgM WB	1 1		Negative Pending				
Other:	/ /	Positive _	_Negative Pending				
A. Why was Lyme diseas	e testing ordered for thi	s patient? <i>Ma</i>	ork all that apply.				
A. Why was Lyme disease testing ordered for this patient? Mark all that apply. Patient had clinical evidence of infection Patient requested Lyme testing							
Patient had exposure to tick habitats Other:							
-			□YES □NO	<u> </u>			
B. Did you prescribe antibiotics for this patient? If yes, indicate type of antibiotic and # of days: YES NO							
Comments:							

Thank you for filling out this form. This information is important to Lyme disease surveillance in West Virginia.

Form B: Patient Lyme Disease Exposure Assessment Tool (2017)

Note: Call patients with erythema migrans (EM)



THIS STEP SHOULD BE LIMITED TO CASES WITH DOCUMENTED EM BY HEALTHCARE PROVIDER

Optional Script

			/
	utside of your home _NO	e county within 30 days of th	e start of your symptoms?
a. If yes, re	eport travel informat	ion:	
Destinat	tion (city, state)	Date of departure (month/day/year)	Date of return (month/day/year)

Thank the patient, and end the call.

Enzyme Immunoassay (EIA) Interpretation Tips

EIA tests detect the amount of antibodies produced by the patient. Immunoglobulin M (IgM), immunoglobulin G (IgG), and combined (or quantitative) IgM/IgG EIA tests are common for Lyme disease diagnostics. □ IgM antibodies are produced by the body <i>early</i> in an infection. The presence of IgG antibodies indicates that the patient was infected with Lyme disease <i>at some point</i> in life.
If the Lyme disease test result you receive has numbers like "0.91" or "5.65," it is an EIA/IFA tes ☐ The higher the number, the more antibodies are being produced.
Some tests will have a reference ranges for "positive," "equivocal," and "negative" test results listed on the laboratory report (see example below).
An "indeterminate" or "equivocal" result means that the level of antibodies detected in the patient's specimen is low. It could also indicate a false positive result. Either way, more information is needed to determine if the patient's immune system produced a response to an infection with Lyme disease; therefore, the EIA and IgM/IgG Western blot are recommended.
"Positive" also means "reactive." "Negative" also means "non-reactive."

Sample reference ranges for LabCorp

≤0.90 Negative 0.91-1.09 Equivocal ≥1.10 Positive

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Interpreting IgG and IgM Western Blots

IgM Western Blot

An IgM immunoblot should be considered positive if two of the following three bands are present:

- -24 kDa (OspC) band
- -39 kDa (BmpA) band
- -41 kDa (Fla) band

Visit the CDC's Lyme disease testing page for more information:

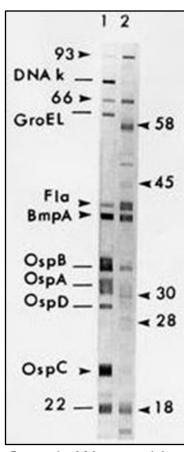
http://www.cdc.gov/lyme/diagnosistesting/index.html

IgG Western Blot

An IgG immunoblot should be considered positive if <u>five of the</u> <u>following ten bands</u>

are present:

- -18 kDa band
- -21 kDa (OspC) band
- -28 kDa band
- -30 kDa band
- -39 kDa (BmpA) band
- -41 kDa (Fla) band
- -45 kDa band
- -58 kDa band
- -66 kDa band
- -93 kDa band

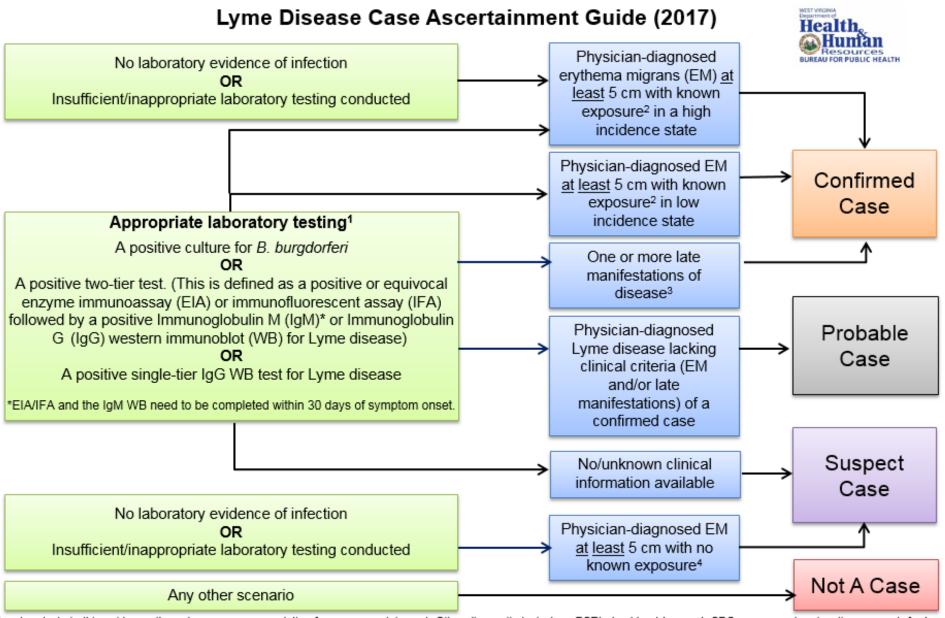


Sample Western blot



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Laboratory tests in this guide are the only ones recommendation for case ascertainment. Other diagnostic tests (e.g. PCR) should not be used. CDC recommends a two-tier approach for Lyme disease testing using serum (EIA/IFA with reflex to Western blot). CSF and synovial fluid are not considered appropriate specimens for two-tier testing.

²Exposure is defined as having been (less than or equal to 30 days before onset of EM) in wooded, brushy, or grassy areas (i.e., potential tick habitats) of Lyme disease vectors. Since infected ticks are not uniformly distributed, a detailed travel history to verify whether exposure occurred in a high or low incidence state is needed. An exposure in a high-incidence state is defined as exposure in a state with an average Lyme disease incidence of at least 10 confirmed cases/ 100,000 for the previous three reporting years. A low-incidence state is defined as a state with a disease incidence of <10 confirmed cases/100,000 (see https://www.cdc.gov/lyme/stats/tables.html). A history of tick bite is not required.

³Late manifestations include musculoskeletal (recurrent, brief attacks of join swelling followed by chronic arthritis), nervous system (lymphocytic meningitis, cranial neuritis, facial palsy (may be bilateral), and radiculoneuropathy, or rarely encephalomyletitis), and cardiovascular (acute onset 2nd -3nd atrioventricular conduction defects that resolve in days to weeks) signs of disease.

4Exposure in a low-incidence state is considered unknown exposure.

PROVIDER QUICKSHEET: LYME DISEASE

JANUARY 2017



IMPORTANT INFORMATION ABOUT SELECTING LABORATORY TESTS

- 1. CDC recommends a two-tier approach for testing serological specimens: IFA/EIA antibody screen, followed by IgM¹ and IgG western blot if IFA/EIA is positive or equivocal.
- 2. Other CDC recommended diagnostic assays for Lyme disease include:
 - A positive culture for B. burgdorferi, OR
 - A positive single-tier IgG² WB test for Lyme disease³.

THE USE OF SINGLE-TIER IGM WESTERN BLOT TESTING IS NOT RECOMMENDED AND WILL NOT BE CONSIDERED CONFIRMATORY FOR PUBLIC HEALTH SURVEILLANCE PURPOSES

RESOURCES FOR PATIENTS

CDC website has several brochures and info sheets for patients: http://www.cdc.gov/lyme/.

RESOURCES FOR HEALTHCARE PROVIDERS

- CDC has a "Resources for Clinicians" page available at: http://www.cdc.gov/lyme/healthcare/clinicians.html
- Information about two-tier testing for Lyme disease is available at: http://www.cdc.gov/lyme/diagnosistesting/LabTest/TwoStep/index.html
- The Infectious Disease Society of America (IDSA) has developed a FREE online CME case study about the diagnosis and management of Lyme disease available at: http://lymecourse.idsociety.org/
- The West Virginia Department of Health and Resources provides information about the state's Lyme disease surveillance system as well as links to useful resources available at: http://www.dhhr.wv.gov/oeps/disease/Zoonosis/Tick/Pages/Lyme.aspx
- The CDC has a "Tickborne Diseases of the United States", reference manual for healthcare providers located at: https://www.cdc.gov/lyme/resources/tickbornediseases.pdf
- The American Academy of Family Physicians (AAFP) provides a diagnostic guideline to aid healthcare providers in diagnosing Lyme disease available at: http://www.aafp.org/afp/2005/0715/p297.pdf

¹ IgM WB is considered positive when at least two of the following three bands are present: 24 kilodalton (kDa) outer surface protein C (OspC)*, 39 kDa basic membrane protein A (BmpA), and 41 kDa (Fla). Disregard IgM results for specimens collected >30 days after symptom onset.

² IgG WB is considered positive when at least five of the following 10 bands are present: 18 kDa, 24 kDa (OspC)*, 28 kDa, 30 kDa, 39 kDa (BmpA), 41 kDa flagellin (Fla), 45 kDa, 58 kDa (not GroEL), 66 kDa, and 93 kDa.

³ While a single IgG WB is adequate for surveillance purposes, a two-tier test is still recommended for patient diagnosis. *Depending upon the assay, OspC could be indicated by a band of 21, 22, 23, 24 or 25 kDA.

